

ACCURACY MEASUREMENT OF TOXIC AND NUTRIENT ELEMENTS IN VEGETAL TISSUE AND SOIL SAMPLES BY USING ISOTOPIC DILUTION ICP-SFMS

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PREPARATION OF REFERENCE MATERIAL

Broccoli

Broccoli was grown by hydroponia, a solution with a mixture of nutrients and toxic metals was used.

PACKAGE
Irradiation (⁶⁰Co)
CRM
168 units for Flower
181 units for Stem

CRUSHING
Particle size (0.5–1) cm

FROZEN (-50 °C)

FREEZE DRY

MILLING

SIEVING
Ro-Tap sieve shaker
Particle size (150 > x > 75) µm

HOMOGENIZATION AND BOTTLING
3-dimensional mixer for 2.5 h

PACKAGE
CRM

Contaminated soil

Sandy soil of rhyolitic origin, from a contaminated site in a mining region of Mexico

COMMUNITING OF MATERIALS
as stone, crushed, milled and pulverized

SIEVING
Ring mills
Tyler Analytical Test Sieves
Ro-Tap Sieve Shaker
Particle size (75–90) µm

HOMOGENIZATION AND BOTTLING
Homogenized in 3-dimensional mixer for 3 h
Rotary cone sample divider and vibratory feeder

PACKAGE
CRM

Irradiation (⁶⁰Co)
512 units of 60 g

PRINCIPLE OF MEASUREMENT

ID-ICPMS Two Steps and Exact Matching Method

The isotope dilution is based on the addition of known amount of an enriched isotope to sample and primary standard. After equilibration of the enriched isotope with natural isotopes, in the sample and in primary standard, the altered isotopic ratios (R_{ix} and R_{iz}), are measured using inductively couple plasma mass spectrometry.

Sample x
 $R_{ix} = x_i/x_x$

Reference z
 $R_{iz} = x_i/x_z$

Enriched isotope, Spike y
 $R_y = x_i/x_y$

Blend x
 $R_{ix} = x_{max}/x_{ix}$

Blend z
 $R_{iz} = x_{max}/x_{iz}$

One Pair

Equipment: Clean room Class ISO 5/7, Double sub-boiling distillation system, Balance Microwave, ICP-SFMS, ICP-QQMS

METHOD SEPARATION

Interferences

¹¹⁰ Cd	¹¹¹ Cd	¹¹² Cd	¹¹³ Cd	¹¹⁴ Cd
⁸⁴ Mo	⁹⁰ Mo	⁹⁶ Mo	⁹⁷ Mo	⁹⁸ Mo
⁸⁴ Zr	⁹⁰ Zr	⁹⁶ Zr	⁹⁷ Zr	⁹⁸ Zr
⁹² Zr	⁹⁴ Zr	⁹⁶ Zr	⁹⁷ Zr	⁹⁸ Zr
⁹² Nb	⁹⁴ Nb	⁹⁶ Nb	⁹⁷ Nb	⁹⁸ Nb
⁹² Mo	⁹⁴ Mo	⁹⁶ Mo	⁹⁷ Mo	⁹⁸ Mo

Mechanism of the ion-exchange separation method

The Cd, Fe, Pb and Zn forms an anionic complexes with Cl⁻, which are stable in HCl (aqueous solution) and the interfering elements do not form stable complexes.

Dowex resin 1X-8
Strong base anion resin, Type 1
Functional group: quaternary amine

Copolymer PS-DVB

The ion exchange chemical reaction is:

$$2 R-N+(R')_3 Cl^- + [CdCl_4]^{2-} \rightarrow [R-N+(R')_3]_2 [CdCl_4]^{2-} + 2 Cl^-$$

$$R-N+(R')_3 Cl^- + [ZnCl_4]^{2-} \rightarrow [R-N+(R')_3]_2 [ZnCl_4]^{2-} + Cl^-$$

$$R-N+(R')_3 Cl^- + [PbCl_4]^{2-} \rightarrow [R-N+(R')_3]_2 [PbCl_4]^{2-} + Cl^-$$

$$R-CH_2N^+(CH_3)_3 Cl^- + [FeCl_4]^{2-} \rightarrow [R-CH_2N^+(CH_3)_3]_2 [FeCl_4]^{2-} + 2 Cl^-$$

Development

Stage 1: Conditioning of resin
Stage 2: Sample loading
Stage 3: Interference elution
Stage 4: Pb, Zn, Cd elution
Stage 5: Washing step

Diagram of species distribution

Separation results

Vegetal tissue, Cd, Pb, Zn
Contaminated soil, Cd, Pb, Zn
Contaminated soil, Fe (HCl 6 M)

For Cd, Pb and Zn:

Stage	1	2	3	4
Conditioning	7% HCl	7% HCl	0.1–2.5% HCl	1% HNO ₃
Sample	2.5% HNO ₃	7% HCl + 3% HClO ₄	0.5–2.5% HNO ₃	2.5% HNO ₃
Elution		7% HCl + 0.5% HClO ₄	12% HCl + 13% HF	
Volume	10–25 mL	3–10 g	10–20 mL	10–15 mL

For Fe:

Stage	1	2	3	4
Conditioning	12 M HCl	6 M HCl	0.5 M HCl	water
Volume	10–25 mL	3–10 g	10–20 mL	10–15 mL

The ion exchange chromatography separation method has been developed for Cd, Fe, Pb and Zn in tissue and soil matrices. Elements as Ca, Na, K, Mg are separated in F1, then the total dissolved solids are reduced. For vegetal tissue and soil, 11 fractions (F) of 4 mL were obtained. The Pb was obtained in F3 and F4, Zn in F6 and Cd in F8-soil, F7-soil. For Fe in soil, 5 fractions (F) of 2, 2, 4, 2 and 2 mL were obtained, the Fe was obtained in F3.

CCQM COMPARISONS

CCQM-K89, Trace and essential elements in Herba Ecliptae

CCQM-K89, Cadmium measurement

CCQM-K89, Zinc measurement

CCQM-K89, Lead measurement

CCQM-K127 "Contaminant and other elements in soil"

CCQM-K127, Cadmium Non-contaminated soil

CCQM-K127, Iron Non-contaminated soil

CCQM-K127, Lead Contaminated soil

CCQM-K127, Cadmium Contaminated soil

CCQM-K127, Iron Non-contaminated soil

CCQM-K127, Lead Non-contaminated soil

For ID-ICPMS two steps and exact matching method, the R_i measured:

Cd: ¹¹¹Cd/¹¹⁴Cd
Fe: ⁵⁷Fe/⁵⁶Fe
Pb: ²⁰⁶Pb/²⁰⁸Pb
Zn: ⁶⁶Zn/⁶⁴Zn

CMC CLAIMS

M. Service Cat. No.	Matrix	Measurand		Dissemination Range of Measurement Capability			Range of Expanded Uncertainties for Measurement Capability			Mechanism(s) for Meas Service Delivery	Source of Traceability	Measurement Technique(s)
		Analyte	Quantity	From	To	Unit	From	To	Unit			
11.1	Vegetal tissue	zinc	Mass fraction	10	80	µg/g	3	4	%	Traceable Reference Materials (MRTC)	CENAM	IC-ID-ICP-SFMS
11.1	Vegetal tissue	calcium	Mass fraction	10	50	mg/g	1.5	4	%	Traceable Reference Materials (MRTC)	CENAM	IC-ID-ICP-SFMS
11.2	Vegetal tissue	cadmium	Mass fraction	1	10	µg/g	1	3	%	Traceable Reference Materials (MRTC), DMR-494a and DMR-494b	CENAM	IC-ID-ICP-SFMS
11.2	Vegetal tissue	lead	Mass fraction	0.5	20	µg/g	5	7	%	Traceable Reference Materials (MRTC), DMR-494a and DMR-494b	CENAM	IC-ID-ICP-SFMS
13.1	Soil	lead	Mass fraction	25	300	µg/g	1.0	1.3	%	PT, DMR-526	CENAM	IC-ID-ICP-SFMS
13.1	soil	cadmium	Mass fraction	0.5	500	µg/g	1.6	1.3	%	PT, DMR-526	CENAM	IC-ID-ICP-SFMS
13.1	soil	Iron	Mass fraction	10	40	mg/g	2.7	1.7	%	PT, DMR-526	CENAM	IC-ID-ICP-SFMS

CONCLUSION

Accuracy measurement method of toxic elements as Cd, Pb and nutrient elements as Fe and Zn in vegetal tissue and soil samples were developed by using ion exchange chromatography separation methods with isotopic dilution ICP-SFMS.

The method were validated by CCQM comparisons CCQM-K89 for vegetal tissue and CCQM-K127 for soil. 7 CMCs has been claimed.

CENAM has been certified three reference materials: 2 batches of Broccoli (Stem and flower) one batch of Contaminated Soil. The range of relative uncertainties is between (1 to 4) %.

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